academic Journals

Vol. 8(43), pp. 5331-5334, 7 November, 2013 DOI: 10.5897/AJAR11.677 ISSN 1991-637X ©2013 Academic Journals http://www.academicjournals.org/AJAR

Full Length Research Paper

Tolerance of fungal and bacterial biocontrol agents to six pesticides commonly used in the control of soil borne plant pathogens

F. A. Mohiddin¹* and M. R. Khan²

¹Seed Technology Research Laboratory, Division of Plant Pathology, SKUAST-K, Srinagar, 190025, India. ²Department of Plant Protection, Aligarh Muslim University, Aligarh, 202 002, India.

Accepted 30 October, 2013

The compatability of fungal (Trichoderma harzianum, Trichoderma virens and Pochonia chlamydosporia) and bacterial biocontrol agents (Bacillus subtilis and Pseudomonas fluorescens) was assessed to 6 pesticides viz., carbendazim, mancozeb, metalaxyl, captan, thiram, and nemacur commonly used by farmers in India for the control of soil borne plant pathogens. The compatability was assessed at different concentrations and the concentration of 60, 1050, 160, 225, 25, and 980 µg/ml of carbendazim, metalaxyl, captan, mancozeb, thiram and nemacur was the safe tolerance limit for T. harzianum whereas the corresponding values for T. virens were 40, 1000, 125, 177, 9, and 700 µg/ml, respectively. The safe tolerance limit for P. chlamydosporia were 37.5 µg carbendazim/ml, 75 µg captan/ml, 100 µg metalaxyl/ml, 5 µg thiram/ml, 110 µg mancozeb/ml and 250 µg nemacur/ml. Among the bacteria, P. fluorescens was found to be more compatible with fungicides than B. subtilis and the maximum tolerance concentration for the former being 2500 µg Thiram/ml, 1600 µg mancozeb/ml, and 50,000 µg/ml for captan and carbendazim. Hence, pesticidal contamination at above concentration in soil will not affect their effectiveness. Moreover, the pesticide tolerance ability broadened the use as these biopesticides in conjugation with pesticides can be applied under integrated disease management for the management of soil borne plant pathogens.

Key words: Biocontrol agents, pesticides, compatability, soil borne plant pathogens.

INTRODUCTION

Soil-borne plant pathogens are highly destructive pathogens and cause tremendous yield losses to all kinds of crops. Control of plant diseases by the use of antagonistic microorganisms can be an effective means (Cook and Baker, 1983). Interaction between biocontrol agents and plant pathogens has been studied extensively and application of biocontrol agents to protect some commercially important crops is promising (Vesseur et al., 1990). A large number of plant diseases have been successfully controlled through fungal and bacterial

antagonists (Sahebani and Hadavi, 2008; Federico et al., 2007; Cook and Baker, 1983; Campbell, 1989; Vidhyasekaran et al., 1997).

Supplementation with specific compounds may provide a competitive advantage for the establishment of the introduced biocontrol agents and improve the biocontrol. In several disease management strategies, the addition of fungicide at reduced rates in combination with biocontrol agents has significantly enhanced disease control, compared to treatments with biocontrol agent

*Corresponding author. E-mail: famohiddin@rediffmail.com.

Treatment	T. harz	zianum	T. vi	rens	P. chlan	iysporia
	MTC*	MIC	MTC	MIC	MTC*	MIC
Carbendazim	60	500	40	405	37.5	250
Metalaxyl	1050	2400	1000	2200	100	500
Captan	160	1000	125	875	75	500
Mancozeb	225	755	177	625	110	350
Thiram	25	150	9	95	5	50
Nemacur	980	1150	700	1000	250	450

Table 1. In vitro compatability of Trichoderma harzianum, T. virens, Pochonia chlamydosporia, with some common pesticides. MTC- Maximum tolerance concentration; MIC- Maximum inhibition concentration.

Each value is mean of 3 replicates; * Concentrations are in (µg/ml).

alone (Frances et al., 2002; Buck, 2004). Integrated use of biocontrol agent with reduced dose of fungicide was effective against fusarium crown and root rot of tomato (Omar et al., 2006), late leaf spot of groundnut (Kishore et al., 2005), rhizoctonia root rot, take-all disease of spring wheat (Duffy, 2000) and post harvest diseases of fruits (Chand-Goyal and Spotts, 1996) compared with the individual components of disease management. The objectives of the present study is to test the growth of different biocontrol agents with commonly used pesticides at different concentrations under *in vitro* conditions for the control of soil borne plant pathogens.

MATERIALS AND METHODS

Six pesticides viz., carbendazim (Bavistin 50 WP), mancozeb (Dithane M-45 75 WP), metalaxyl (Apron 35 SD), captan (Captaf 50 WP), thiram (TMTD 75 WP), and nemacur (Fenamiphos) were tested against the biocontrol agents using poisoned food technique (Grover and Moore, 1961). 50 ml aliquots of PDA (double strength) was taken in an Erlenmeyer flask of 250 ml capacity and sterilized in an autoclave. Different concentrations of the pesticides from 10, 25, 50, 125, 250, 500, 1000, 2000, 3000, and 5000 µg/ml were prepared in distilled water. 50 ml of a concentration was aseptically transferred to the Erlenmeyer flask containing 50 ml PDA. 5 Petri plates (90 mm diameter) for each concentration of the fungicides were prepared by pouring 20 ml PDA aliguots in each plate and allowed to solidify. Thereafter, the plates were seeded centrally with a 3 mm disc of 4 days old culture of Trichoderma harzianum, Trichoderma virens and Pochonia chlamydosporia. PDA plates without a fungicide but inoculated with the fungi served as a control. The inoculated plates were incubated at 25 ± 2°C for 5 days. The radial growth of the colony in each treatment was measured and the percent inhibition of growth was calculated by the formula:

I = C - T/C x 100

Where, I = Percent growth inhibition; C = Radial growth in control (mm); T = Radial growth in treated plates (mm). And ED_{90} (maximum inhibition concentration) and ED_{50} (safe tolerance concentration) were determined. To determine the compatability of *B. subtilis* and *P. fluorescens* with same pesticides, 25 to 50,000 µg/ml concentrations were prepared in double distilled water. Double strength nutrient agar was used as medium for both the bacteria. 20 ml of nutrient agar containing desired concentration was poured in Petri plates and left over night to observe contamination, if any. Thereafter, 0.1 ml of overnight cultures was

spread over the solidified plates with a glass spreader. The plates were incubated at $30 \pm 2^{\circ}$ C for 24 h and bacterial colonies were identified and counted.

RESULTS

The maximum growth of *T. harzianum* was inhibited at a concentration of 500, 2400, 1000, 755, 150, and 1150 μ g/ml of carbendazim, metalaxyl, captan, mancozeb, thiram, and nemacur, respectively.

The corresponding value for *T. virens* were 405, 2200, 875, 625, 95 and 1000 μ g/ml of of carbendazim, metalaxyl, captan, mancozeb, thiram and nemacur, respectively (Table 1).

The fungicides at concentrations of 60 μg carbendazim/ml, 1050 µg metalaxly/ml, 160 μg captan/ml, 225 µg mancozeb/ml, 25 µg Thiram/ml, and 980 µg Nemacur/ml seem to be safe tolerance limit for T. harzianum. For T. virens the safe tolerance limit were 40 carbendazim/ml, 125 µg captan/ml, 177 μg μg mancozeb/ml 1000 µg metalaxyl/ml, and 700 μq nemacur/ml (Table 1). Pochonia chlamydosporia showed less tolerance to the 6 pesticides tested.

The fungus was inhibited by the concentrations of 250 μ g carbendazim/ml, 500 μ g each of captan and metalaxyl/ml, 350 μ g mancozeb/ml, 50 μ g thiram/ml and 450 μ g nemacur/ml. Whereas, the safe tolerance limit for *P. chlamydosporia* were 37.5 μ g carbendazim/ml, 75 μ g captan/ml, 100 μ g metalaxyl/ml, 5 μ g thiram/ml, 110 μ g mancozeb/ml, and 250 μ g nemacur/ml (Table 1). Biocontrol bacteria were found more tolerant to fungicides than fungi (Table 2).

The maximum tolerance concentration for *Bacillus subtilis* were 3200 μ g captan/ml, 60 μ g thiram/ml 600 μ g mancozeb/ml. Whereas, in case of carbendazim, the bacteria showed tolerance even for a concentration of 50,000 μ g/ml (Table 2).

Pseudomonas fluorescens was found to be more compatible than *B. subtilis* with fungicides, the maximum tolerance limit for the former being 2500 μ g Thiram/ml, 1600 μ g mancozeb/ml and 5 μ g/100 ml for captan and carbendazim and 8000 μ g nemacur/ml (Table 2).

Treatment	B. su	btilis	P. fluorescens		
	MTC*	MIC	MTC	MIC	
Carbendazim	50, 000	-	50, 000	-	
Metalaxyl	7,000	10,000	10,000	25,000	
Captan	3200	4000	50, 000	-	
Mancozeb	600	1000	1600	2000	
Thiram	60	100	2500	3000	
Nemacur	3500	4200	8000	9000	

Table 2. In vitro compatability of Bacillus subtilis and Pseudomonas fluorescens with some common pesticides.

Each value is mean of three replicates; *Concentrations are in (μ g/ml); MTC, Maximum tolerance concentration; MIC, Maximum inhibition concentration.

DISCUSSION

The compatability tests revealed that the Trichoderma species showed more tolerance to metalaxyl as compared to other pesticides used in the study. Similar results have been obtained by other workers. Sharma et al. (2001) found that, T. harzianum is showing more tolerance to metalaxyl as compared to carbendazim. More or less similar results have been found by other workers also (Nallathambi et al., 2009; Mukhopadhyay et al., 1986; Mukherjee et al., 1989, Papavizas et al., 1982; Viji et al., 1997). Different workers have reported chlorothalonil, captan and captafol as tolerant for T. harzianum even at higher concentrations up to 2000 mg/ml in spore germination tests (Abdel-Moity et al., 1982; Papavizas et al., 1982; Mishra et al., 2004). The biocontrol bacteria viz., Pseudomonas fluorescens and Bacillus subtilis were found more tolerant to fungicides than fungi. This may be due to the reason that, some bacteria can use pesticides as nutrients and hence can tolerate higher concentrations of chemicals (Kishore and Jacob, 1987; Aislabie and Jones, 1995).

The present study clearly demonstrated that, soil borne plant pathogens can be successfully managed by combined application of biocontrol agents with a cheap fungicide like carbendazim, mancozeb, metalaxyl, captan, thiram, and nemacur commonly used by farmers in India at low doses. Also pesticidal contamination in soil will not affect the biocontrol agent effectiveness and hence can be easily applied in conjugation with the pesticides for the control of soil borne plant pathogens.

REFERENCES

- Abdel-Moity H, Papavizas GC, Shatia MN (1982). Induction of new isolates of *Trichoderma harzianum* tolerant to fungicides and their experimental use for control of white rot of onion. Phytopathology 72:396-400.
- Aislabie J, Jones LG (1995). A review of bacterial degradation of pesticides. Austr. J. Soil Res. 33:925-942.
- Buck JW (2004). Combination of fungicides with phylloplane yeasts for improved control of *Botrytis cinerea* on geranium seedlings. Phytopathology 94:196-202.

Campbell R (1989). Biological Control of Microbial Plant Pathogens. Cambridge University Press, Cambridge.

- Chand-Goyal T, Spotts RA (1996). Postharvest biological control of blue mold of apple and brown rot of sweet cherry by natural saprophytic yeasts alone or in combination with low doses of fungicides. Biol. Control 6:253-259.
- Cook RJ, Baker KF (1983). The Nature and Practice of Biological Control of Plant Pathogens. American Phytopathological Society, St. Paul, MN.
- Duffy B (2000). Combination of pencycuron and *Pseudomonas fluorescens* strain 2-79 for integrated control of *Rhizoctonia* root rot and take-all of spring wheat. Crop Protect. 19:21-25.
- Federico GR, Maria MR, Marcela F, Sofía NC, Adriana MT (2007). Biological control by *Trichoderma* species of *Fusarium solani* causing peanut brown root rot under field conditions. Crop Protect. 26:549-555.
- Frances J, Vilardell P, Bonaterra A, Badosa E, Mantesinos E (2002). Combination of *Pseudomonas fluorescens* EPS288 and reduced fungicide dose for control of *Penicillium* rot during post harvest storage of pear. Acta Hortic. 596:883-886.
- Grover RK, Moore DJ (1961). Adaptation of *Sclerotinia fructicola* and *Sclerotinia laxa* to higher concentrations of fungicides. Phytopathology 51:399-401.
- Kishore GM, Jacob GS (1987). Degradation of glyphosate by *Pseudomonas* sp. PG2982 via a sarcosine intermediate. Journal of Biological Chemistry 262: 12164-12168.
- Kishore GK, Pande S, Podile AR (2005). Management of late leaf spot of groundnut with chlorothalonil-tolerant isolates of *Pseudomonas* aeruginosa. Plant Pathol. 54:401-408.
- Mishra A, Sharma SD, Patel SI (2004). Cross tolerance of *Trichoderma harzianum* Rifai to fungicides. Indian Phytopathol. 38:207-211.
- Mukherjee PK, Upadhyay JP, Mukhopadhyay AN (1989). Biological control of *Pythium* damping off of cauliflower by *T. harzianum*. J. Biol. Control 3:119-124.
- Mukhopadhyay AN, Brahamabhatt A, Patel GJ (1986). *Trichoderma harzianum* a potential biocontrol agent of tobacco damping-off. Tobacco Res. 12:26-35.
- Nallathambi P, Umamaheswari C, Thakore BBL, More TA (2009). Postharvest management of ber (*Ziziphus mauritiana* Lamk) fruit rot (*Alternaria alternata* Fr. Keissler) using *Trichoderma* species, fungicides and their combinations. Crop Protect. 28:525-532.
- Omar I, O'Neill, TM, Rossall S (2006). Biological control of Fusarium crown and root rot of tomato with antagonistic bacteria and integrated control when combined with the fungicide carbendazim. Plant Pathol. 55:92-99.
- Papavizas GC, Lewis JA, Abdel Moity TH (1982). Evaluation of new biotypes of *T. harzianum* for tolerance to benomyl and enhanced biocontrol capabilities. Phytopathology 72:126-132.
- Sahebani N, Hadavi N (2008). Biological control of the rootknotnematode Meloidogyne javanica by Trichoderma harzianum. Soil Biol. Biochem. 40:2016-2020.
- Sharma SD, Mishra A, Pandey RN, Patel SJ (2001). Sensitivity of *Trichoderma harzianum* to Fungicides. J. Mycol. Plant

Pathol. 31:251-253.

- Vesseur V, Arigoni F, Anderson H, Defago G, Bompeix G, Seng JM (1990). Isolation and characterization of *Aphanocladium album* chitinase over producing mutants. J. General Microbiol. 136:2561-2567.
- Vidhyasekaran P, Rabindran R, Muthamilan M, Nayar K, Rajappan K, Subramanian N, Vasumathi K (1997). Development of powder formulation of *Pseudomonas fluorescens* for control of rice blast. Plant Pathol. 46:291-297.
- Viji G, Manibhushan R, Baby UI (1997). Non target effect of systemic fungicides on antagonism microflora of *R. solani*. Indian Phytopathol. 50:324-328.